

Lab 7 Simulation of DNA hybridizations

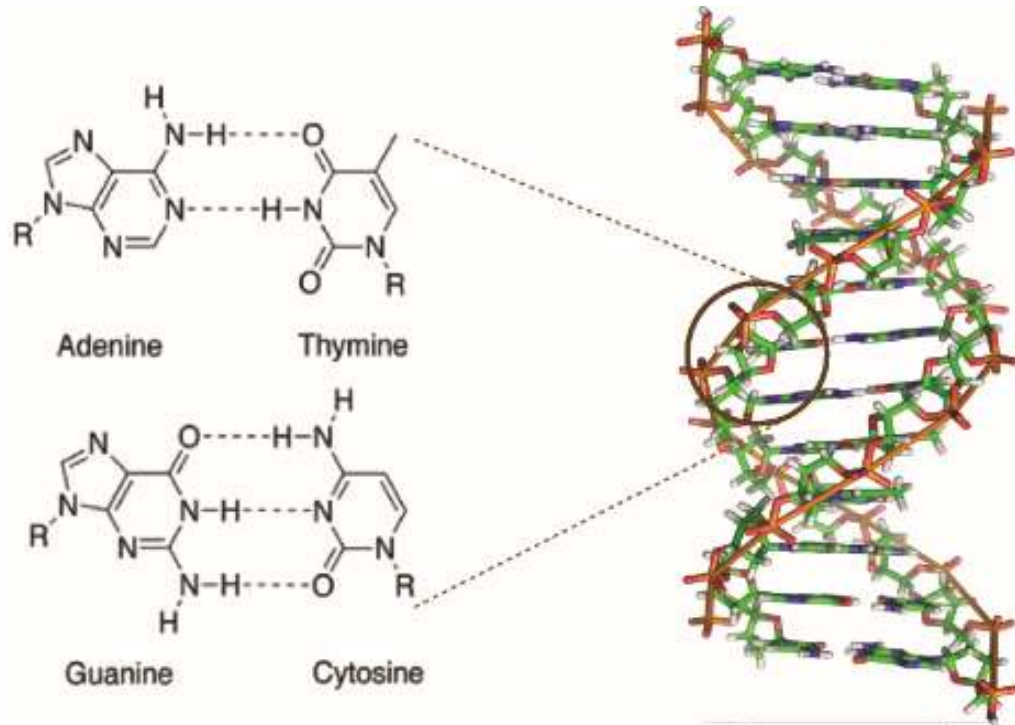
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DNA hybridization energy calculation



ssDNA: single-stranded

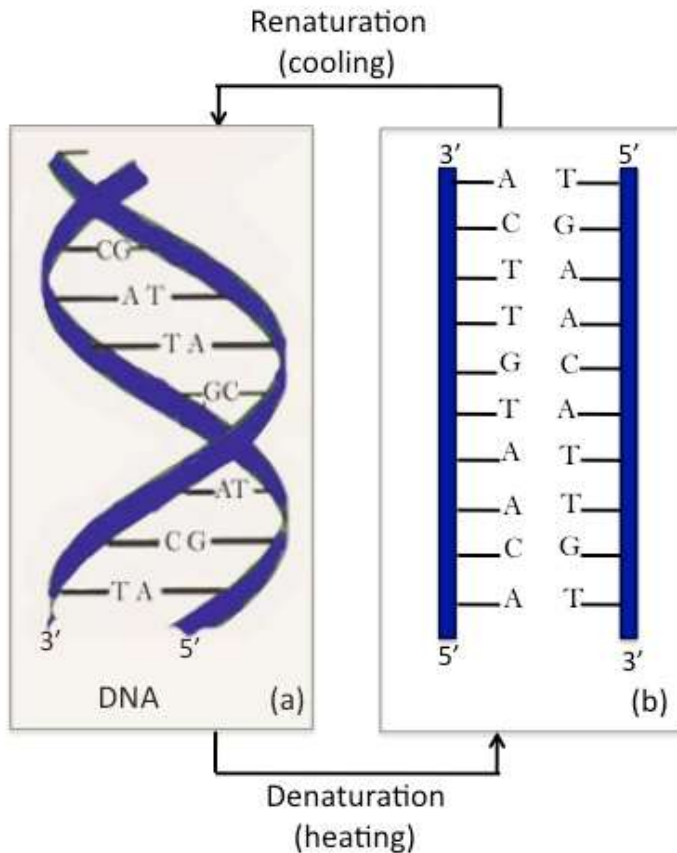
dsDNA: double-stranded

- dsDNA is stabilized by base pairing
- Hybridization energy follows a thermal physical reaction process:

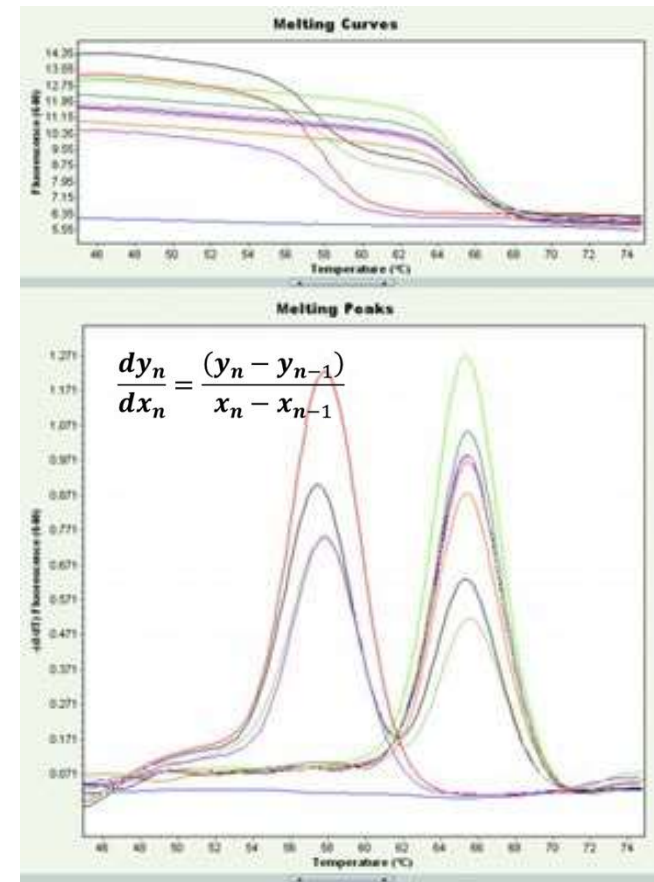
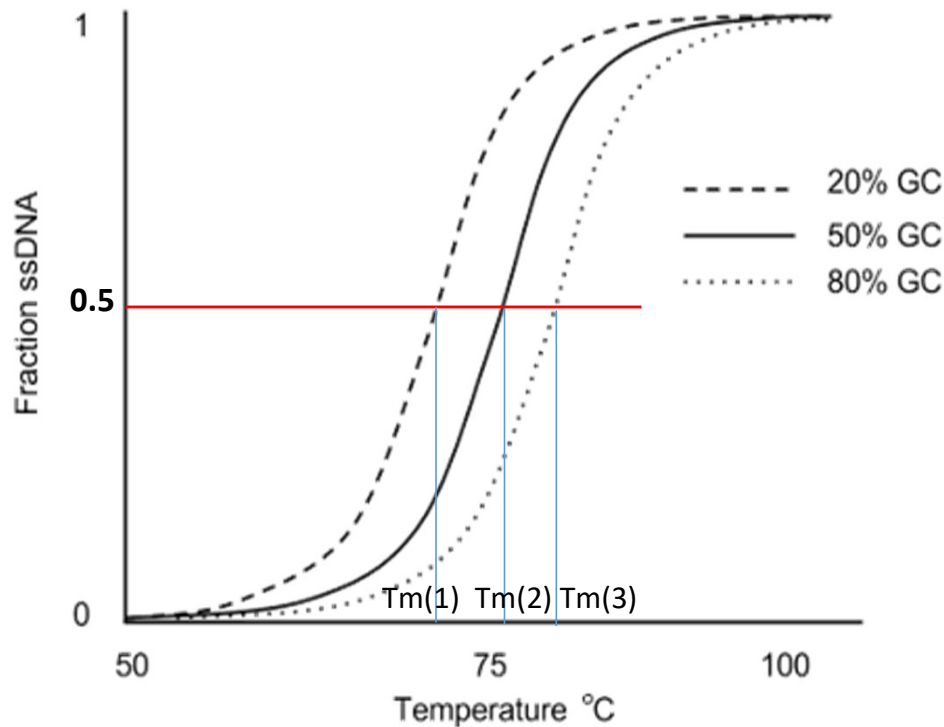
$$\Delta G(\text{total}) = \sum \Delta G (\text{individual base pairs}) + \sum \pi - \pi \text{ stacking} - \sum \text{charge repulsion}$$

- Melting temperature is an important parameter for DNA hybridization stability

DNA thermal melting (or thermal denaturation): The dsDNA complexes may be dissociated by thermal denaturation, which is referred to as thermal melting.



- Melting temperature (T_m): the melting temperature of DNA refers to the temperature at which 50% of DNA in a sample has denatured from double-stranded DNA (dsDNA) to single-stranded DNA (ssDNA). Sensitive measurement of the melting curve of a sample of DNA can be used to detect single nucleotide differences between two DNA samples.
- Other melting methods: Chemical reagent can also disrupt dsDNA, such as urea, SDS, detergent ... It is called chemical denaturation



Melting Temperature (T_m) for dsDNA complexes with different GC%. Note, a dsDNA with **high GC%** shows a higher melting temperature due to the increased thermal stability. In addition, **magnesium ions** also play a key role to stabilize dsDNA by shielding the negative charges of phosphate backbone.

T_m , practically, is calculated by slope derivative analysis, that T_m is the temperature at the peak slope value. Generally, the first-order derivative analysis can be used to produce the slope curve and identify T_m value

Lab Goal:

- **Learn NUPACK to simulate DNA hybridizations;**
- **Learn the fitting of DNA melting temperature;**
- **Understand the how DNA melting temperature is impacted by GC%, length, and Mg²⁺ conc.**

DNA computation and simulation with NUPACK

The screenshot displays the NUPACK web interface with the following components:

- Navigation Bar:** Analysis (selected), Design, Utilities, Download, and user profile icons.
- Sub-navigation:** Input (selected), Intro, Demos.
- Material:** RNA (selected), DNA.
- Temperature:** 37.0 °C, with a Melt toggle switch.
- Model Options:** Tube: Tube 1.
- Tube 1 Configuration:**
 - Tube: Tube 1
 - Species: A table with columns for Strand, Sequence, and Concentration.
- Complexes:** Max complex size: 1 strands.

| Strand | Sequence | Concentration |
|----------|----------|---------------|
| Strand 1 | | 1.0 μM |

• Set Parameters for NUPACK

- Can analyze DNA and RNA;
- Set temperature and Melting analysis
- Run user-defined sequences and concentration
- In the “More Option”: Set Salt concentrations (Na⁺ and Mg²⁺)

The screenshot displays the NUPACK web interface with the following settings:

- Material:** DNA (selected), RNA (unselected). Temperature: 37.0 °C. Melt analysis is enabled (toggle switch).
- Model Options:**
 - Parameters:** dna04
 - Ensemble:** All stacking
 - Salts:** Na⁺ 1.0 M, Mg²⁺ 0.0 M
- Tube:** Tube 1
- Species:**

| Strand | Sequence | Concentration | |
|----------|----------|---------------|---|
| Strand 1 | | 1.0 μM | ✕ |

+ Add Strand

Task 1 Use Nupack to simulate and analyze dsDNA melting graph

- **Set Melting Temp analysis for poly (A-T) 20: Choose “DNA” and “Melt”**
- Set Min 25 C and Max 85 C, step 2 C
- Set “dna04” and “All stacking”; set Na⁺ to 0.137 M, Mg⁺⁺ to 0 M
- Set name of “Tube” as: poly (A-T)20
- Set two strands of poly A20 and polyT20. You can use “Add strand” to add the second strand. Conc. = 1 uM

The screenshot shows the Nupack web interface with the following configuration:



- Material:** DNA (selected), RNA (unselected)
- Temperature:** Melt (toggle on), Min: 25 °C, Step: 2 °C, Max: 85 °C
- Model Options:**
 - Parameters:** DNA, dna04 (NUPACK3)
 - Ensemble:** All stacking
 - Salts:** Na⁺ 0.137 M, Mg⁺⁺ 0.0 M
- Tube:** poly (A-T)20
- Species:**


| Strand | Sequence | Concentration | |
|-----------|-----------------------|---------------|---|
| poly A 20 | aaaaaaaaaaaaaaaaaaaaa | 1.0 μM | × |
| poly T 20 | tttttttttttttttttttt | 1.0 μM | × |



+ Add Strand

Task 1 Use Nupack to simulate and analyze dsDNA melting graph


- Set “Max Complex Size” to: 2 strands for dsDNA
- Click “Analyze”

Tube   View Ensemble

▼ Species 

| Strand | Sequence | Concentration | |
|-------------|---|---------------|---|
| poly A 20 ▼ | aaaaaaaaaaaaaaaaaaaaa  | 1.0 μM ▼ | ✕ |
| poly T 20 ▼ | ttttttttttttttttttttt  | 1.0 μM ▼ | ✕ |

[+ Add Strand](#)

Complexes 

Max complex size

strands

▸ Include or exclude specific complexes

[+ Add Tube](#)

[Analyze](#)

Task 1 Use Nupack to simulate and analyze dsDNA melting graph

- Click results from analysis view melting graph.
- You can drag the temperature bar to view the hybridization changes at a specific temperature



Task 1 Use Nupack to simulate and analyze dsDNA melting graph

- Download data from NUPACK simulation
- Click “Download” to save the simulated DNA hybridization data in excel.



| A | B | C |
|----------------|----------------------------|---|
| Temperature(C) | Fraction of bases unpaired | |
| 25 | 0.083582 | |
| 27 | 0.088888 | |
| 29 | 0.095704 | |
| 31 | 0.103196 | |
| 33 | 0.11521 | |
| 35 | 0.124953 | |
| 37 | 0.136696 | |
| 39 | 0.151554 | |
| 41 | 0.171085 | |
| 43 | 0.197727 | |
| 45 | 0.235084 | |
| 47 | 0.287373 | |
| 49 | 0.359036 | |
| 51 | 0.452491 | |
| 53 | 0.564164 | |
| 55 | 0.681804 | |
| 57 | 0.787051 | |
| 59 | 0.865391 | |
| 61 | 0.914699 | |

• Fitting of “Melt Temp” from downloaded data

- Tm fitting with IC50 online tool: <http://www.ic50.tk/index.html>
- Copy and paste data into the blank box, click submit. IC50 is the Melt Temp of poly (A-T)20

Paste in X, Y values:

```
25    0.083582479
27    0.088888131
29    0.095704074
31    0.103195933
33    0.115209652
35    0.124953308
37    0.136696167
39    0.151554048
41    0.171084897
43    0.197727253
45    0.235084209
47    0.287372781
49    0.359035691
51    0.452490502
```

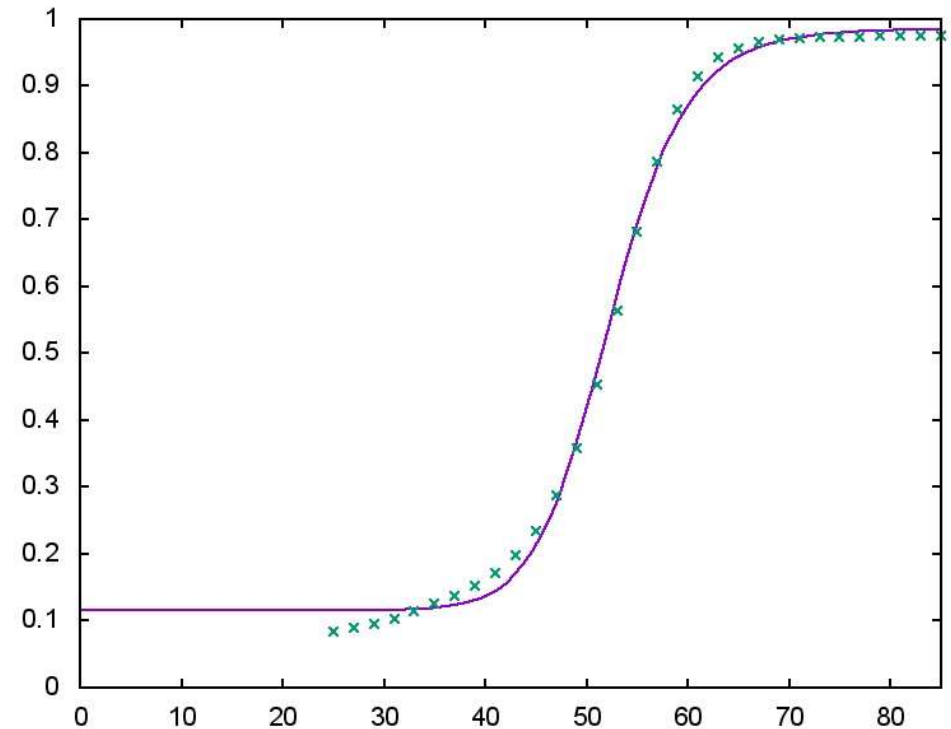
Submit

Tick here and re-submit if curve-fit is bad.

Right-click on image to save.

Results:

| | |
|------------------|---------------------------------|
| Minimum | 0.115961 +/- 0.006163 (5.315%) |
| Maximum | 0.986135 +/- 0.006087 (0.6172%) |
| IC ₅₀ | 52.3513 +/- 0.1722 (0.329%) |
| Hill coeff. | 13.7464 +/- 0.5523 (4.018%) |



Task 1 Use Nupack to simulate and analyze dsDNA melting graph
(from 25 –85 C, 3 C per step)

- **GC%:** Thermal melting graph for 1 uM poly(A-T)₂₀ and poly (C-G)₂₀; at 0.137M Na⁺, 0 Mg²⁺, Fitting for T_m
- **Length:** Thermal melting graph for 1 uM poly(A-T)₂₀ and 1 uM poly(A-T)₄₀; at 0.137 M Na⁺, 0 Mg²⁺, Fitting for T_m
- **Mg²⁺:** Thermal melting graph for 1 uM poly(A-T)₂₀ at 0.137 M Na⁺, 0 M Mg²⁺, Fitting for T_m.

Do it again for salt conditions at 0.137 M Na⁺, 0.01 M Mg²⁺.

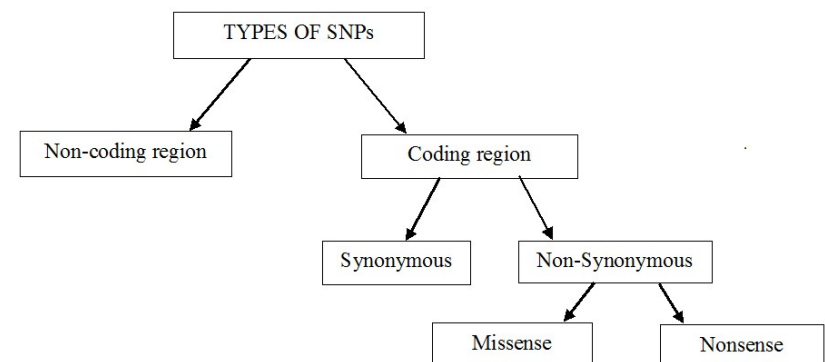
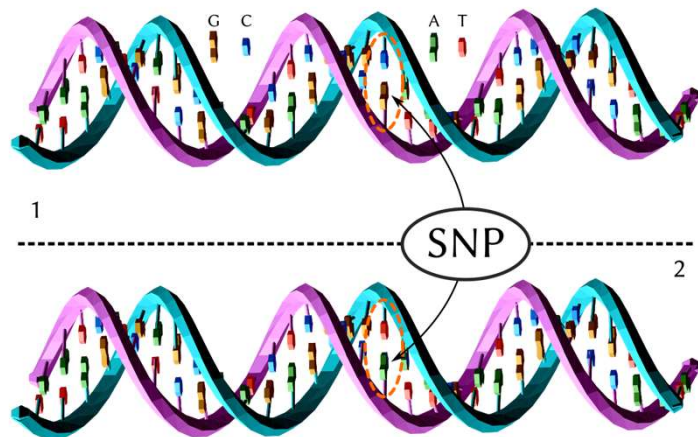
- How to conclude the parameters that affect the melt Temp of dsDNA?

Task 2. DNA hybridization melting temperature to detect SNP/SNV (Single-nucleotide Polymorphism/Variants)

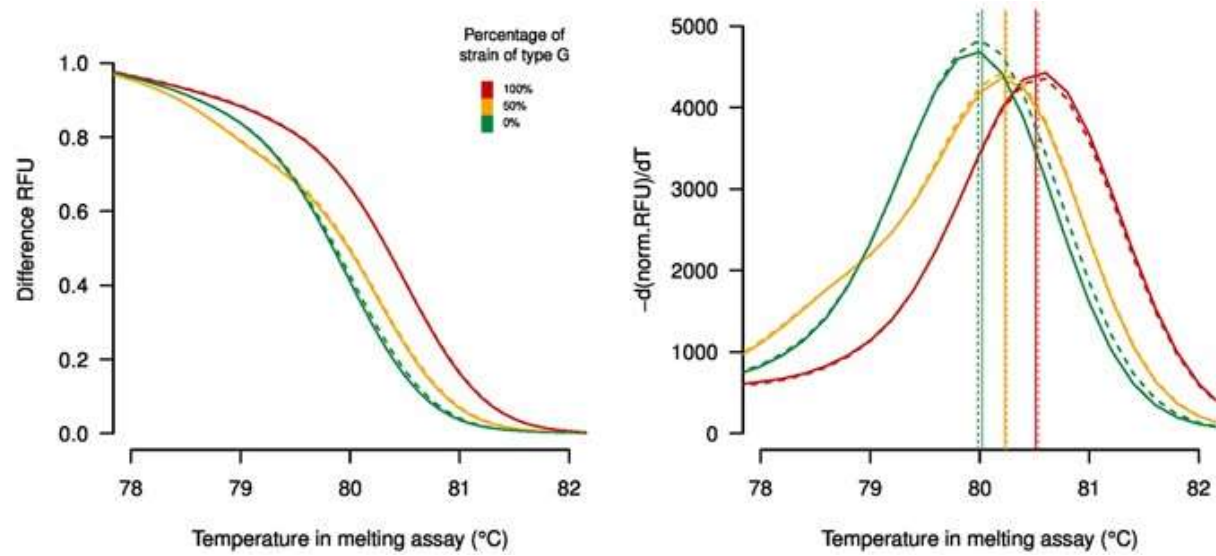
A single nucleotide polymorphism, or SNP (pronounced "snip"), is a variation at a single position in a DNA sequence among individuals. For example, at a specific base position in the human genome, the G nucleotide may appear in most individuals, but in a minority of individuals, the position is occupied by an A. This means that there is a SNP at this specific position, and the two possible nucleotide variations – G or A – are said to be the alleles for this specific position. Although a particular SNP may not cause a disorder, some SNPs are associated with certain diseases.

SNP may or may not cause an amino acid mutation.

Single-nucleotide variant (SNV) is a variation in a single nucleotide. SNVs differ from SNPs in that a SNV can be somatic[9] and **can be caused by cancer**,[10] but a SNP has to segregate in a species' population of organisms. SNVs also commonly arise in molecular diagnostics such as designing PCR primers to detect viruses, in which the viral RNA or DNA sample may contain SNVs.



SNP/SNV detection by PCR with melting temperature difference, **first-order derivative analysis**

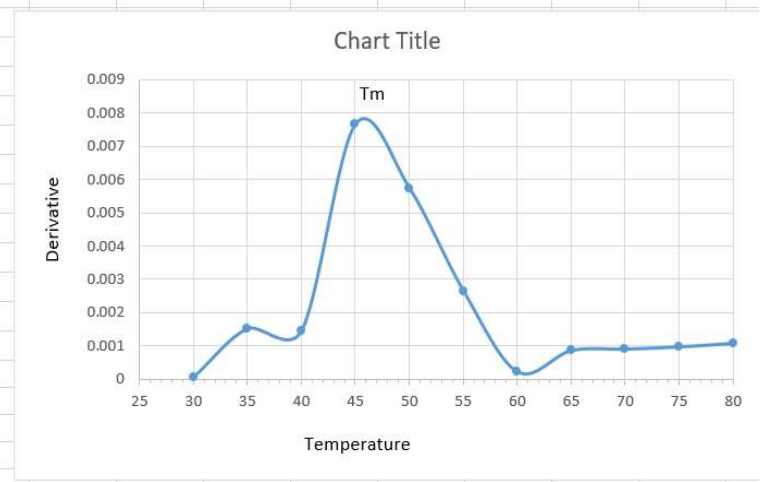


Derivative Analysis with Excel

$$\frac{dy_n}{dx_n} = \frac{(y_n - y_{n-1})}{x_n - x_{n-1}}$$

| Temperature | A260 | Derivative |
|-------------|---------|-------------------|
| 25 | 1.12084 | |
| 30 | 1.12119 | = (B3-B2)/(A3-A2) |
| 35 | 1.12882 | 0.001526403 |
| 40 | 1.13616 | 0.001466918 |
| 45 | 1.17456 | 0.00768013 |
| 50 | 1.2033 | 0.00574801 |
| 55 | 1.21659 | 0.002658391 |
| 60 | 1.21771 | 0.000224924 |
| 65 | 1.22205 | 0.00086658 |
| 70 | 1.22659 | 0.000908089 |
| 75 | 1.23146 | 0.000974798 |
| 80 | 1.23686 | 0.001080463 |

Tm ~ 46 C



Tm is the temperature with the highest derivative value

Example: CYP1A2 SNP

rs762551, also known as -164A>C or -163C>A, is a SNP encoding the CYP1A2*1F allele of the CYP1A2 gene. The CYP1A2 gene encodes a member of the cytochrome p450 family of proteins, which metabolize nutrients and drugs. One well known substrate of CYP1A2 is caffeine; individuals who carry one or more CYP1A2*1C alleles are "slow" caffeine metabolizers, whereas carriers of the variant CYP1A2*1F are "fast" caffeine metabolizers.

| Primer | Sequences |
|-----------------------|--|
| TaqMan probe – TET | 5'-CTC TGT GGG CCC AGG ACG CAT-3' |
| TaqMan probe – FAM | 5'-TC TGT GGG CAC AGG ACG CAT GG-3' |
| TaqMan Forward primer | 5'-TTT CCA GCT CTC AGA TTC TGT GAT-3' |
| TaqMan Reverse primer | 5'-GGA TAC CAG AAA GAC TAA GCT CCA TC-3' |
| Forward primer* | 5'-TTC CCC ATT TTG GAG TGG TC-3' |
| Reverse primer* | 5'-CCG AGA AGG GAA CAG ACT GG-3' |

Nordmark et. al. 2002

[Home](#) > [Search Tool](#) > [Search Results](#) > [C__8881221_40](#)

SNP ID: rs762551
Gene ▼CYP1A2
Gene Name cytochrome P450 family 1 subfamily A member 2
Set Membership: > HapMap > DME > Validated > Inventoried
Chromosome Location: -
Polymorphism: C/A, Transversion Substitution
Context Sequence [VIC/FAM]: TGCTCAAAGGGTGAGCTCTGTGGGC[C/A]CAGGACGCATGGTA
GATGGAGCTTA

Task 2: Use Nupack to predict thermal melting graphs and Tm at 0.137 M Na+ and 0 Mg++

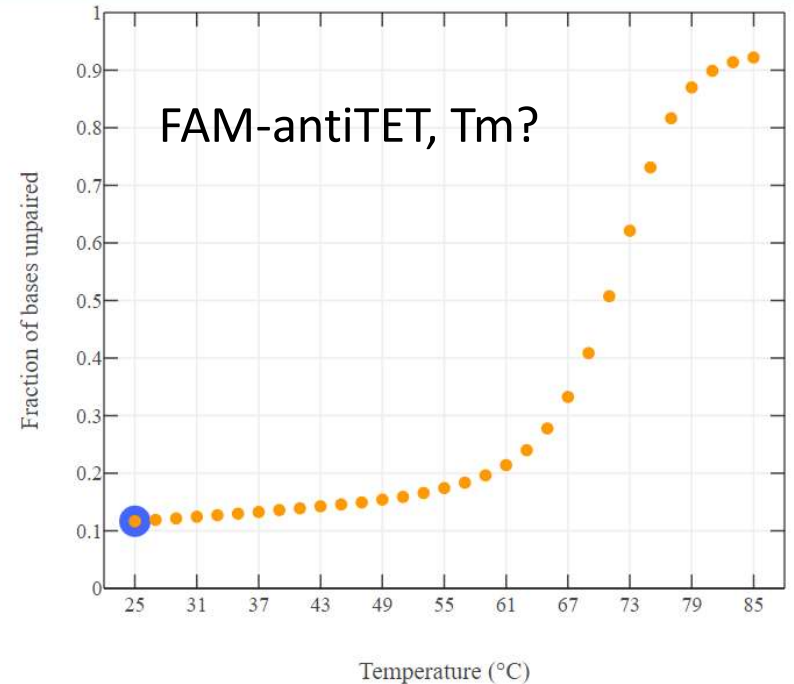
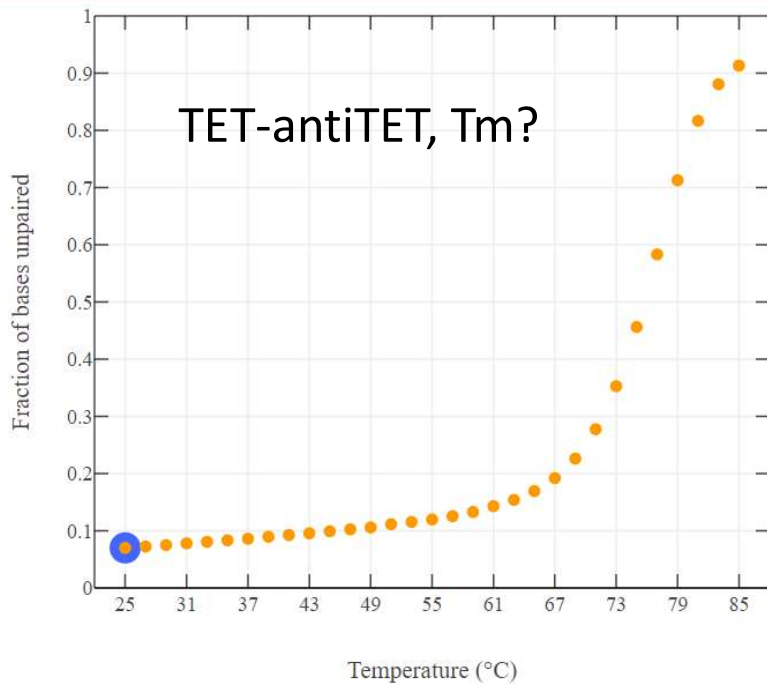
1) Anti-TET + TET seq (1 uM) and anti-TET + FAM seq (1 uM)

| | Sequence | Tm (C) |
|------------|---|--------|
| Target TET | CTCTGTGGGCC C CAGGACGCAT | |
| Target FAM | CTCTGTGGGGC A CAGGACGCAT | |
| Anti-TET | A TGCGTCCTG G GCC C ACAGAG | |

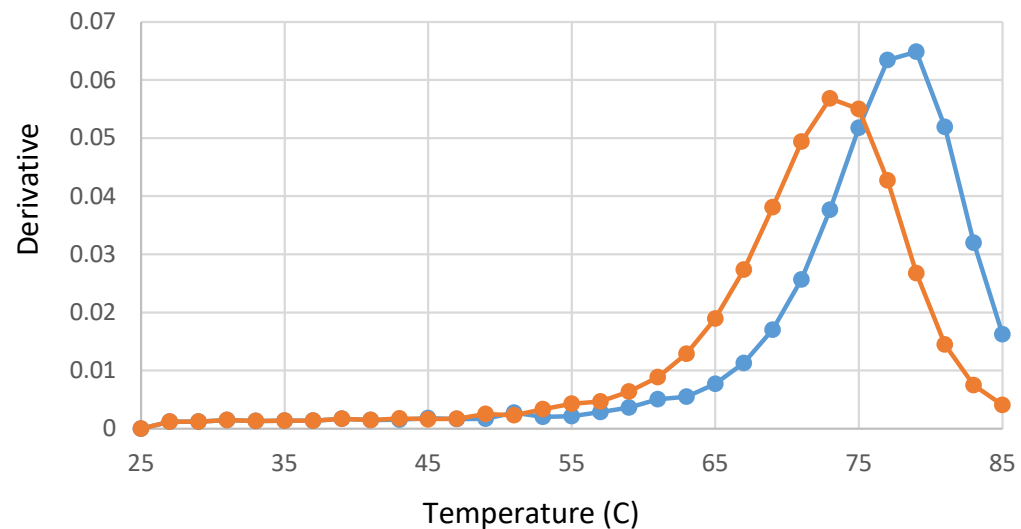
2) Based on Tm difference, suggest a **temperature** that produces the biggest difference for hybridizations between TET/anti-TET and FAM/anti-TET

| | Sequence | Test Temperature (C) | Hybridization yield (%) |
|------------|-------------------------------------|----------------------|-------------------------|
| Target TET | CTCTGTGGGGC C CAG GACGCAT | | |
| Target FAM | CTCTGTGGGGC A CA GGACGCAT | | |

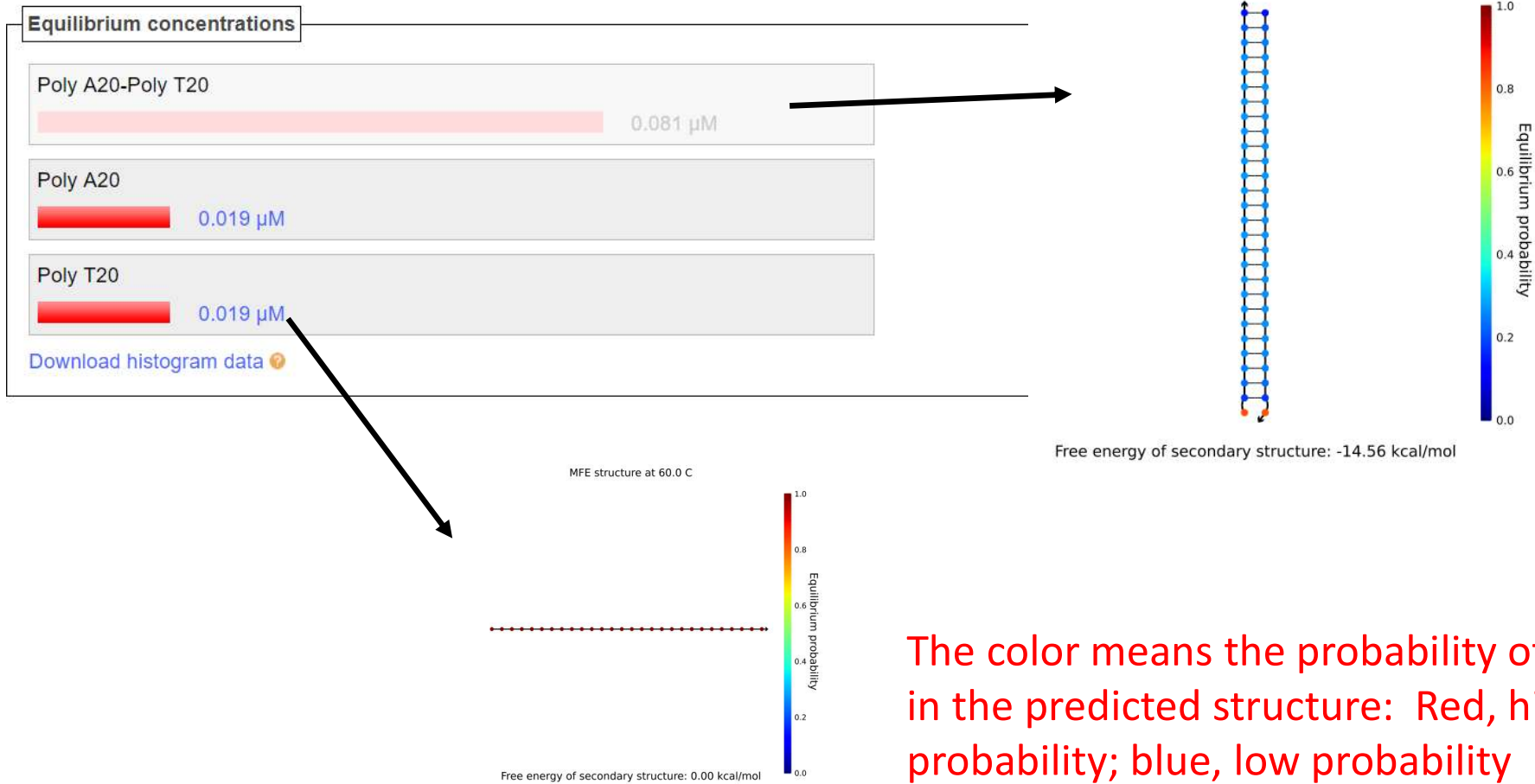
• Melt Temp by first-order derivative analysis



First-order derivative analysis



Hybridization Yield%



The color means the probability of the base in the predicted structure: Red, high probability; blue, low probability

Nupack can predict the equilibrium conc. of hybridizations. You can calculate the product yield:

e.g.

$$[\text{Target dsDNA}]/[\text{total ssDNA input}] = 0.081/0.1 = 0.81=81\%$$

Task 3. Predict the secondary structure of RNA (RNA mode at 37 C) : the folding structure of SARS-COV-2 (cause COVID-19) amplicons for PCR diagnosis

N1 :

GATAATGGACCCCAAATCAGCGAAATGCACCCCGCATTACGTTTGGTGGACCCTCAGATTCAACTGG
CAGTAACCAGAAT

N2:

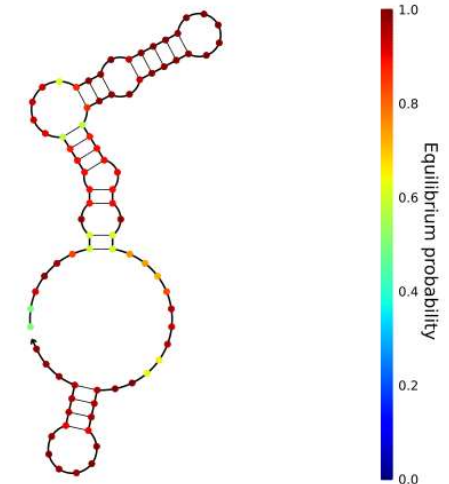
ACTAATCAGACAAGGAACTGATTACAAACATTGGCCGCAAATTGCACAATTTGCCCCAGCGCTTCAG
CGTTCTTCGGAATGTCGCGC

N3:

AGACGGCATCATATGGGTTGCAACTGAGGGAGCCTTGAATACACCAAAGATCACATTGGCACCCGC
AATCCTGCTAACAATGCTGCAATCGTGCTACAACCTCCTCAAGGAACAACA



The color means the probability of the base in the predicted structure: Red, high probability; blue, low probability

MFE structure at 37.0 C



Free energy of secondary structure: -19.40 kcal/mol




- Calculation mode for RNA
- Under RNA mode, you cannot choose salt concentration
- Set Temp to 37 C, 1 strand

Material  Temperature:  Melt


RNA DNA


37 °C

▼ Model Options


Parameters  Ensemble  Salts 



RNA rna06 All stacking Na⁺ 0.137 M Mg⁺⁺ 0 M

▼ Tube: N1 


Tube  [View Ensemble](#)

N1

▼ Species 

| Strand | Sequence | Concentration |
|--------|---|---|
| N1 | N1 : GAUAAUGGACCCCAAAAUCAGCGAAAUGCACCCCGCAUUACGUUUGGUGGACCC  | 1 μM <input type="text"/>  |

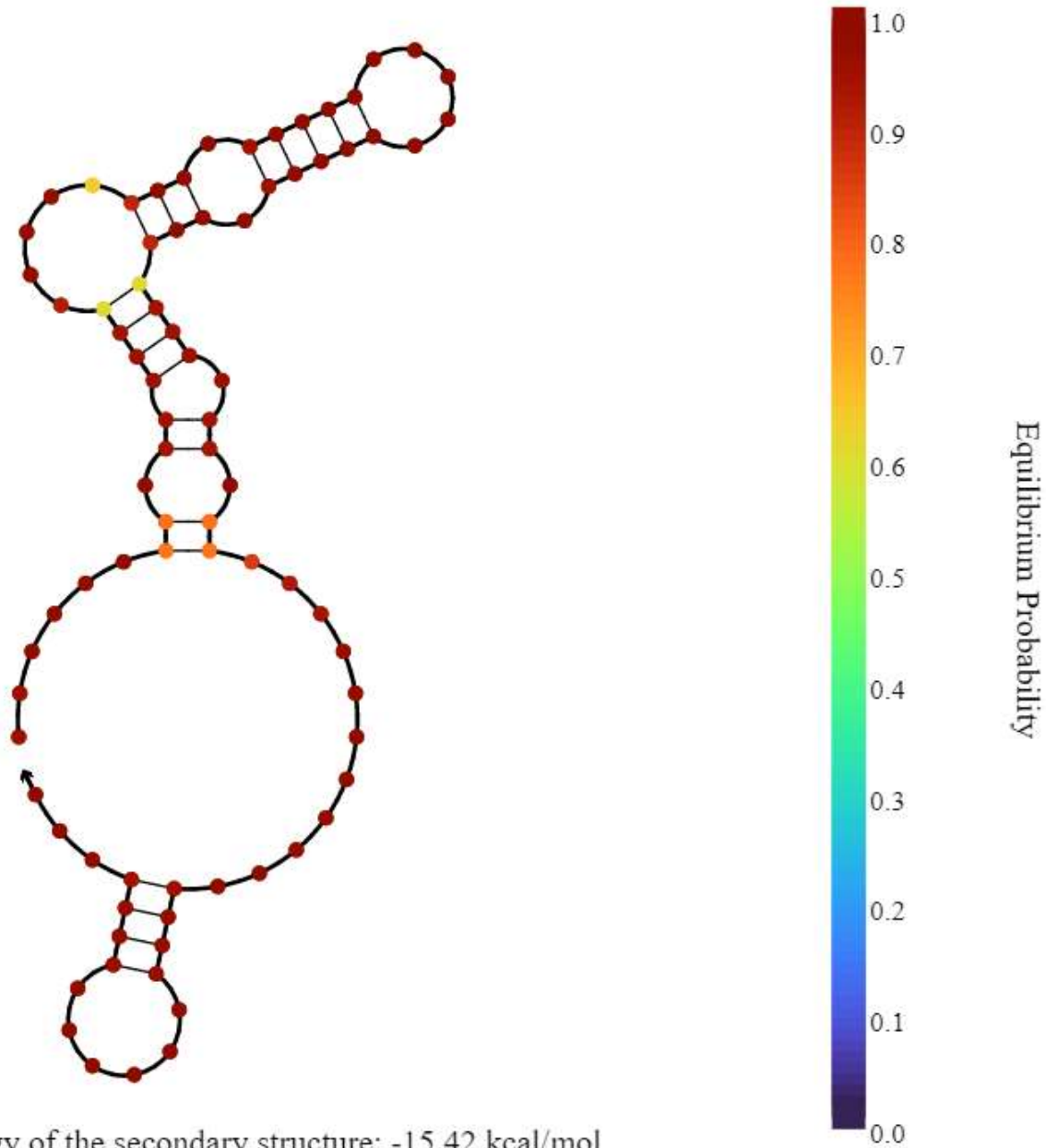
[+ Add Strand](#)

Complexes 

Max complex size

1 strands

MFE proxy structure at 37°C



Data sheet

Task 1 (at least 2 C interval, from 20 – 80 C) Na⁺ fixed at 0.137 M

- Figure 1 Thermal melting curves for poly(A-T)₂₀ vs poly (C-G)₂₀
- Figure 2 Thermal melting curves for poly(A-T)₂₀ vs poly (A-T)₄₀
- Figure 3 Thermal melting curves for poly (A-T)₂₀ at 0 Mg²⁺ vs 0.01 M Mg²⁺
- T_m value fitting

Task 2

Use salt condition: 0.137 M Na⁺, 0 Mg

Thermal melting curves for TET and FAM, T_m value fitting using **1st order of derivative analysis**

Pick up a temperature based on derivative analysis , compare hybridization yield difference between TET and FAM at this fixed temperature.

Task 3

- Folding structure for N1, N2, N3 amplicons